

Kooman, J. P., Dekker, M. J., Usvyat, L. A., Kotanko, P., van der Sande, F. M., Schalkwijk, C. G., Shiels, P. G. and Stenvinkel, P. (2017) Inflammation and premature aging in advanced chronic kidney disease. *American Journal of Physiology: Renal Physiology*, 313(4), F938-F950.  
(doi:[10.1152/ajprenal.00256.2017](https://doi.org/10.1152/ajprenal.00256.2017))

This is the author's final accepted version.

There may be differences between this version and the published version. You are advised to consult the publisher's version if you wish to cite from it.

<http://eprints.gla.ac.uk/144603/>

Deposited on: 08 August 2017

**Title: Inflammation and premature aging in advanced chronic kidney disease**

**Authors: Jeroen Kooman<sup>1</sup>, Marijke Dekker<sup>1</sup>, Len Usvyat<sup>2</sup>, Peter Kotanko<sup>3,4</sup>, Frank van der Sande<sup>2</sup>, Casper Schalkwijk<sup>1</sup>, Paul G Shiels<sup>5</sup>, Peter Stenvinkel<sup>6</sup>**

**Affiliation of the authors:**

<sup>1</sup> Maastricht University Medical Center, Maastricht, Netherlands

<sup>2</sup> Fresenius Medical Care North America, Waltham, MA, USA

<sup>3</sup> Renal Research Institute, New York, NY, USA

<sup>4</sup> Icahn School of Medicine at Mount Sinai, New York, NY, USA

<sup>5</sup> Institute of Cancer Sciences, MVLS, University of Glasgow, Glasgow, UK

<sup>6</sup> Division of Renal Medicine M99, Dept of Clinical Science Technology and Intervention, Karolinska Institutet, Stockholm, Sweden

**Word count Main Body:**

5287

**Word count Abstract:**

220

**Corresponding author:**

Jeroen Kooman,

[Jeroen.kooman@mumc.nl](mailto:Jeroen.kooman@mumc.nl)

Department of Internal Medicine, division of Nephrology

University Hospital Maastricht, The Netherlands

*Abstract*

Systemic inflammation in end-stage renal disease (ESRD) is an established risk factor for mortality and a catalyst for other complications which are related to a premature aging phenotype, including muscle wasting, vascular calcification and other forms of premature vascular disease, depression, osteoporosis and frailty. Uremic inflammation is also mechanistically related to mechanisms involved in the aging process, such as telomere shortening, mitochondrial dysfunction, and altered nutrient sensing, which can have direct effect on cellular and tissue function. In addition to uremia-specific causes such as abnormalities in the phosphate- Klotho axis, there are remarkable similarities between the pathophysiology of uremic inflammation and so-called “inflammaging” in the general population. Potentially relevant, but still somewhat unexplored in this respect are abnormal or misplaced protein structures as well as abnormalities in tissue homeostasis, which evoke danger signals through damage associated molecular patterns (DAMPs) as well as the senescence associated secretory phenotype (SASP). Systemic inflammation, in combination with the loss of kidney function, can impair the resilience of the body to external and internal stressors by reduced functional and structural tissue reserve, and by impairing normal organ crosstalk, thus providing an explanation for the greatly increased risk of homeostatic breakdown in this population. In this review, the relation between uremic inflammation and a premature aging phenotype, as well as potential causes and consequences are discussed.

## 54 *Introduction*

55 End-stage renal disease (ESRD) is characterized by a greatly increased risk of cardiovascular and  
56 infectious mortality, as well as by structural and functional abnormalities of various organ systems,  
57 most notably the cardiovascular, the immune, and the musculoskeletal system. Substantial  
58 similarities in phenotype exist between ESRD and the aging process. About 30-50% of pre-dialysis,  
59 hemodialysis (HD), and peritoneal dialysis (PD) patients have serologic evidence of an active  
60 inflammatory response that is related to adverse outcomes (17, 18, 132). Persistent “uremic<sup>i</sup>  
61 inflammation”, as this phenomenon has been coined in the literature (148), resembles that observed  
62 in various chronic diseases as well as in the aging process in the general population (“inflammaging”).  
63 (68, 137).

64 Although several reviews already have addressed the causes and nature of uremic  
65 inflammation in detail (18, 60, 132), recent findings have revealed novel causes and mechanisms of  
66 uremic inflammation as well as the catalytic role of systemic inflammation changing the risk factor  
67 profile. Since systemic inflammation may be both a cause and consequence of (premature) aging this  
68 may be of relevance for the marked discrepancy between chronological and biological age observed  
69 ESRD (68, 137). The aim of this review is to discuss potential similarities between the  
70 pathophysiology of inflammaging and systemic uremic inflammation, as well as on the putative  
71 relation between uremic inflammation and premature aging.

72

## 73 **Mechanisms of uremic inflammation**

### 74 *Premature aging of the immune system*

75 The immune system is a complex orchestration of cells, cytokines and other molecules that act in a  
76 paracrine, autocrine, or endocrine manner to protect the human organism primarily against  
77 infectious disease (114). Whereas this mechanisms is essential for survival, when chronically

stimulated, it can become maladaptive and is in this sense an example of antagonistic pleiotropy (142). In the uremic milieu, abnormalities in the immune response are characterized by an abnormal activation *and* a reduced functioning of components of the innate and adaptive immune system, which contributes to systemic inflammation and increased susceptibility for infectious complications (58). Various abnormalities, such as an impaired neutrophilic phagocytic capacity, depletion of B-cells and naïve T-cells as well as depletion of dendritic cells contribute to reduced functioning of the immune system (“immunosenescence”) (8, 58, 155). Important similarities exist between the effects of aging and ESRD on the adaptive immune response (8, 9), whereas a comparable systemic activation of the innate immune response may also be observed during aging (“inflammaging”) (68). Both factors argue for a premature aging process of the uremic immune system (9).

#### *Activation of the innate immune system*

The activation of the *innate* immune system in uremia is characterized by an increase in pro-inflammatory cytokines, such as TNF and interleukin (IL)-6. Activation of transmembranous Toll-like receptors (TLR4), classically by pathogen-associated molecular patterns (PAMPS), induces transcription factors, such as nuclear factor- $\kappa$ B (NF $\kappa$ B) (89, 96), which is a master regulator of cytokine secretion. Moreover, IL-6 stimulates hepatic C-reactive protein (CRP) production (28). Importantly, NF $\kappa$ B is also upregulated by oxidative stress, and can be stimulated by cytokines, such as TNF, leading to self-stimulation of the inflammatory process (116) [Figure 1].

NLPR (NACHT, LRR and PYD domains-containing protein) inflammasomes form another class of pattern recognition receptors (PRR). These lead to upregulation of IL-1B and IL-18 expression through caspase 1. Inflammasomes are intracellular protein complexes, which are activated by a variety of triggers, including cytokines, reactive oxygen species (ROS) as well and damage-associated molecular patterns (DAMPS) (76) [Figure 1]. An increase in NLRP3 mRNA expression, as well as upregulation of caspase 1, IL-1B and IL-18 was observed in peripheral blood mononuclear cells of HD patients

compared to controls (42). Whereas circulating myeloid cells and M1 macrophages are the primary effector cells of uremic inflammation (42), the inflammatory response can also be triggered in other cell types, such as vascular endothelium and vascular smooth muscle cells (14, 42, 89, 147).

#### *Defective regulation of the inflammatory process*

The inflammatory process is, under physiological circumstances, meticulously regulated, with an intricate balance between pro- and anti-inflammatory parameters (135). For the regulation of innate immune system, the sirtuin family, and most notably Sirtuin-1, plays an important role, modulated by Nf-kB inhibition through different pathways, such as AMPK, PGC-1 $\alpha$  and PPAR (160). Sirtuin-1 down-regulation may also lead to an imbalance between M1 pro-inflammatory and M2 anti-inflammatory macrophages in favor of the former. Sirtuin-1 downregulation has been observed in aging and metabolic syndrome and relates to inflammatory markers (59). Reduced sirtuin 3 expression also relates to mitochondrial damage and increased oxidative stress in animal models of acute kidney injury (102). Noteworthy in this context are recent observations indicating that at least two miRNAs (hsa-mir-217 and hsa-mir-125b) regulate sirtuin and AKT activity, as well as the mTOR pathways involved in regulating aging processes across taxa (91), providing a biochemical link between cellular ageing, stress and damage responses. Although its role in the pathogenesis of uremic inflammation needs to be established hsa-miR-125b is a critical component of a range of immunological phenomena, including host-defense responses, autoimmunity, immune cell differentiation and IL-4 and INF- $\gamma$  expression (145). A study using genome-wide gene expression profiling identified a differential expression of 80 genes between 10 hemodialysis (HD) patients and controls; variations of these genes are linked to pro-inflammatory pathways, such as the TLR pathways. Using interaction network analysis, 68 differentially expressed miRNA were connected to 47 genes suggesting an important role for miRNA in the regulation of uremic inflammation (170).

## **Arguments for a premature aging process in ESRD in relation to systemic inflammation**

The first argument for an uremic premature aging process is the increase in age-adjusted mortality, which is an aspecific marker of ageing. A recent editorial argued against the indiscriminate use of the term premature aging and proposed four domains of the aging phenotype (87) i.e. 1) changes in body composition, 2) impaired energy balance, 3) impaired homeostatic mechanisms and 4) neurodegeneration. A reduced lean tissue mass mass and an increase in fat mass (sarcopenic obesity) have been reported in ESRD (85, 86); both relate to the expression of inflammatory markers (50). A low bone density is another prevalent feature of ESRD that relate to inflammation and adverse outcomes (22). Regarding energy balance, both maximum aerobic exercise as well as tissue glucose uptake are reduced in CKD (20, 153). While energetic efficiency appears to be reduced, resting energy expenditure are increased in ESRD, in relation to inflammation (158). Also, there is an inverse relation between physical activity (or physical capacity) with inflammatory markers (33).

Except from anemia with high erythropoietin, the impaired homeostatic mechanisms mentioned by Margolick and Ferrucci (87) are all prevalent in uremic inflammation (68, 137). Notably, in keeping with these feautres, neurodegeneration, impaired cognitive function and balance are already prevalent in earlier stages of CKD (44, 88), whereas brain atrophy is a well known complications of ESRD (32). Next to these four domains, vascular progeria is a common finding in the inflamed uremic phenotype and significant associations between vascular calcification and increased vascular stiffness with inflammatory biomarkers are often reported (68). Thus, according to the phenotypic criteria, it can be concluded that an argument for the presence of a premature aging syndrome can be well made. Moreover, recent studies found that abnormalities in the kidney and blood vessels in patients with renal failure were associated with a progeric and senescent phenotype (138, 143).

## **Mechanistic relations between uremic inflammation and (premature) aging**

The next question is whether uremic inflammation is mechanistically related to biological ageing (174). For this purposes, a reflection on the relation between uremic inflammation and aging hallmarks is relevant (80). In non-uremic mice, chronic inflammation, induced by the knockout of the NFκB subunit 1, resulted in telomere shortening and a phenotype of progressive aging (56). In dialysis patients, increased *telomere attrition* was observed in comparison to age-matched controls and related to inflammatory markers (19, 24, 68). *Oxidative stress*, generally regarded as a major contributor to biological ageing, is increased in ESRD and reciprocally related to (uremic) inflammation (140, 172). Uremic inflammation impairs *nutrient sensing*, which is also considered an important hallmark of aging (80). TNF and IL-6 induce catabolism by stimulation of the ubiquitin proteasome complex and blunt anabolic pathways by IGF resistance and aberrant mTOR regulation (39, 68, 135). These effects, which can be considered a cellular stress response, can explained both by a direct effect of inflammation on these pathways. An alternative explanation is reduced energy availability to the cell because of shifting of energy to the inflammatory response and a concomitant increase in sympathetic nervous system activity (142). Moreover, systemic inflammation is also related to a *decrease in endothelial progenitor cells* in uremic patients (49). This might play a role in impaired vascular repair, although in the same study no link between endothelial progenitor cells and endothelial dysfunction was observed (106). A recent study by Kramann et al (70) show that critical adventitial progenitors (Gli1+ cells) may be relevant therapeutic targets for mitigation of vascular calcification. Senescence may also make the cell more susceptible to damage evoked by uremic toxins and or oxidative stress (16).

## **Causes of uremic inflammation**

### *Abnormalities in mineral metabolism*

Abnormalities in mineral metabolism appear to be another important link in the relation between inflammation and progeria (75). In adenine-induced CKD rats, dietary phosphate increased systemic TNF as well as tissue (e.g. in kidney heart and aorta) mRNA expression in a dose dependent matter,



which was prevented by the use of the phosphate binder lanthanum carbonate (166). In CKD4 patients the phosphate binder sevelamer increased fetuin-A, which is a negative acute phase protein and an inhibitor of extracellular matrix mineralization) (139) (45) (15). The mechanisms behind phosphate-induced inflammation may at least be partly dependent upon generation of oxidative stress and activation of NFκB (175). Phosphate may also lead to osteoblast induction of vascular smooth muscle cells (VSMC), which might subsequently release inflammatory mediators especially in combination with a senescent phenotype (7). Indeed, increased serum phosphate levels may drive cellular and physiological senescence (73). A surprising result was observed in a study in uremic rats, where the calcification process of dietary phosphate was actually enhanced by a very low protein diet, and was also associated with systemic inflammation, as evidenced by an increase in TNF levels and a decline in fetuin levels (167). Another proof linking phosphate to progeria is a recent study that reports that inorganic phosphate activates the mTOR pathway (57).

Fetuin mediates the formation of calciprotein particles (CPP), circulating colloidal complexes containing calcium and phosphate, which are catabolized by the mononuclear phagocytic system (130) and might lead to a reduction of mineral stress. However, formation of CPP also results in the reduction of circulating and intracellular fetuin levels, with a potential loss of protection against the extracellular calcification and to the transformation of VSMC (128). The calcification propensity of serum, which is inversely reflected by the “maturation time” ( $T_{50}$ ) of CPPs, of serum was related to all-cause mortality in patients with CKD stages 3-4 as well as in renal transplant recipients (61, 129). It can be speculated that when formation of CPP exceeds clearance, the cytotoxic CPP induce pro-inflammatory cytokines (130).

#### *Defective anti-aging mechanisms*

An intriguing relation appears to exist between uremic inflammation and the anti-aging protein Klotho (105). The anti-aging properties of Klotho in endothelial cells were explained by inhibition of NFκB translocation from cytoplasm to the nucleus by stabilisation of the NFκB /IKK complex, which

protected these cells from senescence (13). Klotho expression was reduced by TNF, TWEAK and NFκB activation (101, 142). Next to this epigenetic repression of Klotho gene expression via accumulation of protein bound toxins may be operative (144). As Klotho is also a potent inhibitor of vascular calcification, a self-reinforcing interaction between uremic inflammation, phosphate accumulation, decreased Klotho expression, cellular senescence, and vascular calcification may be operative in the uremic milieu (51).

### *Gut dysbiosis*

The causes of inflammation specifically related to dialysis treatment, such as vascular access, bioincompatibility of dialysis membranes contamination of dialysis solutions or the use of intravenous iron, have been summarized extensively in previous reviews (17, 34, 40) [Figure 2]. The same holds true for potentially modifiable factors, such as periodontitis (17, 40, 71, 72). An emerging factor with relevance for both inflammaging and uremic inflammation is gut dysbiosis (83) (110, 156). Shi et al. (125) observed bacterial DNA in 12 out of 52 ESRD patients and a correlation with CRP and IL-6 levels. Elevated endotoxin levels, which are related to bacterial DNA (125), have also been observed in uremic plasma (93) and soluble CD14 predicts mortality in HD patients (109). Moreover, the microbial metabolite Trimethylamine-N-oxide (TMAO), which has been linked to adverse cardiovascular outcome, correlates with uremic inflammation and is an independent predictor of mortality in CKD (99). Although the origin of the increased endotoxin levels in uremia remain to be elucidated it is likely that a translocation of gut microbiome due to increased gut permeability is the primary contributor. Constituents of tight junctions like claudin-1, occludin and ZO-1 were reduced in the colon of uremic rats (157). A recent study showed that depletion of tight junction proteins coincided with a reduction in nuclear factor erythroid 2-related factor 2 (Nrf2), which has a central role in the regulation of intracellular oxidative stress (74). Recently, a study studied the interaction of gut dysbiosis, aging and inflammation. In wild-type mice, the microbial constitution of the faeces, changed with aging whereas gut permeability increased, leading to translocation of bacterial

products into the blood and induction of systemic inflammation. Remarkably, these age-related changes were absent in TNF- $\alpha$  deficient mice, which was explained by an interaction between the inflammatory state of the host and the intestinal microbiome (150)

#### *Regulation of oxidative stress*

Uremic toxins such as phosphate, protein bound toxins and advanced glycation end (AGEs) products, can evoke inflammatory pathways directly or mediated by oxidative stress (47). ROS stimulates the inflammatory process through NF $\kappa$ B signaling (78). As the uremic milieu may down-regulate Nrf2 (107), which inhibits NF $\kappa$ B and upregulates a large number anti-oxidative genes (90), impaired Nrf2 activity likely contributes to uremic inflammation. Perturbed expression of these expression factors also appears to contribute to senescence(176).

#### *Non-enzymatic glycation*

During the ageing process, increased protein damage takes place as a result of non-enzymatic glycation (108). Protein glycation was viewed originally as a post-translational modification of proteins that accumulated slowly on extracellular and long-lived proteins throughout life. In the extracellular matrix, so called advanced glycation endproducts (AGEs) caused aberrant cross-linking resulting in a decrease of elasticity in vessels leading to arterial stiffness and hypertension, i.e. hallmarks of vascular ageing. The physiological consequences of the formation of AGEs in the aetiology of a range of important age-related diseases, such as ESRD, have been described (82). In addition to the slow formation of AGES, glycation adducts are also formed in a fast manner on cellular and short-lived extracellular proteins and on DNA. The highly reactive methylglyoxal (MG) is a key compound involved in the very fast generation of glycation adducts on proteins, lipids and DNA. Methylglyoxal is mainly generated as a by-product of glycolysis. To counteract the deleterious effects of MG, organisms contain an enzymatic glyoxalase defense system comprised of glyoxalase I (GLO1) and GLO2, in which MG is converted to D-lactate. GLO1 is a key enzyme in regulating the levels of

MGO and AGEs. It has been shown that GLO1 and GLO2 activity decreases in human arterial tissues and red blood cells during the aging process (63, 94). The downstream consequences of GLO1 reduction have been demonstrated by an overexpression of the GLO1 homologue in *C. Elegans*, resulting in an increase of the mean and maximum lifespan by ca 30%; silencing the GLO1 homologue decreased the lifespan by about 50% (100, 122). Thus, since the balance between the production of MGO and its detoxification by GLO1 contribute to the ageing process, managing this balance is important for the prevention of age-related health problems (164). Next to their direct effects on the (vascular) aging process, AGEs can also induce inflammation via NFkB activation and subsequent expression of pro-inflammatory cytokines (141) in target cells, such as VSMC. A relation between serum pentosidine levels and monocyte activation markers was observed in CKD (162). On the other hand, blockade of the RAGE receptor reduced oxidative stress and atherosclerosis in uremic mice, but not the mRNA expression of inflammatory mediators in aortic smooth muscle cells (11). AGEs could also contribute to inflammation by endoplasmatic reticulum (ER) stress (90), which occurs when the demand for protein folding, a major task of the ER, exceeds capacity (31). ER stress may induce inflammation and cellular senescence by Nfkb activation and increased translocation of  $Ca^{2+}$  into the cytosol (31, 79, 117, 123). It has also been demonstrated that uremic serum induces ER stress in human umbilical vein endothelial cells (HUVEC), via NFkB upregulation (171).

#### *Danger associated molecular patterns (DAMPS)*

An important factor in the pathogenesis of inflammaging with potential relevance for uremic inflammation is the presence of misplaced or misfolded molecules, which serve as so-called danger associated molecular patterns (DAMPS), which are non-microbial inducers of inflammation that are evolutionary strongly preserved. DAMPS signal cellular and tissue stress and might evoke an inflammatory response by TLRs, RAGE and/or inflammasomes (38). Various DAMPS have been identified with portential relevance for CKD, such as extracellular ATP, uric acid, S100 proteins and the high mobility group box 1 HMBG1 protein (77, 120). Whereas there is accumulating evidence for a role of DAMPS in the pathogenesis of localized inflammation in CKD (113), the evidence for a role

of DAMPS in the pathogenesis of systemic uremic inflammation is yet limited. However, an inverse relation between renal function and serum levels of HMBG1 (12) and a relation between serum levels of HMBG1 and TNF, IL-6 and CRP (177) have been reported.

Accumulation of DAMPS may be related to a disturbance in *autophagy* (38, 77). Autophagy serves to remove damaged intracellular organelles and to enable the recirculation of essential nutrients. Complex interactions exist between inflammation and autophagy, which may act as a double ended sword for the individual. On one hand, autophagy may eliminate inflammatory triggers by removal of DAMPS. On the other hand, whereas systemic inflammation may induce autophagy through a cellular stress response, autophagy may also release DAMPS and, thus, induce inflammation (77, 118, 119). Similar to oxidative stress and inflammation, autophagy may be beneficial for cellular survival during short-term or minor insults, but have detrimental effects during prolonged or excessive activation. Reduced autophagy was observed in uremic leukocytes (21). However, autophagy of phosphate loaded VSMC was found to be protective against vascular calcification (25). Conversely, in an experimental model of renal failure, inflammation markers were related to increased autophagy in muscle (159). Thus it is not yet clear if increased or defective autophagy is a causative factor in uremic inflammation (161).

#### *Cellular senescence*

A factor which is considered highly important in the pathogenesis of inflammaging is the *senescence-associated secretory phenotype* (SASP), in which senescent cells release pro-inflammatory cytokines such as TNF, IL-1, IL-6 and IL-8 (23) (112), which poison the surrounding tissues. The inflammatory process can progress from the cell to the tissue and whole body environment by extracellular spillover and through what is termed the communicome or secretome, which can act at local, tissue, as well as systemic levels. In this communicome, circulating cytokines, miRNA and extracellular vesicles may be involved (38). Senescent mononuclear cells with increased expression of pro-

inflammatory cytokines were observed in HD-patients treated with cellulosic membranes, but not in serum of predialysis patients (111). Additionally, phosphate and indoxyl sulphate induce VSMC senescence (103, 165). VSMC in human carotid plaques of non-uremic patients showed evidence of a SASP accompanied by secretion of IL-1a (41). Studies on the SASP in CKD are limited. Although it has been suggested that the SASP is involved in the pathogenesis of chronic allograft nephropathy (131) the role of SASP in the pathogenesis of the uremic phenotype needs to be addressed further. We recently showed that severe uremic arterial calcification was associated with increased vascular expression of *CDKN2A*<sup>p16<sup>INK4a</sup></sup>, increased number p<sup>16</sup> positive cells and SASP (138). Notably, in an epidemiological cohort, <10% of the variability in IL-6 expression in the circulation could be explained on the basis of cellular ageing, expressed by telomere length (127). A recent study showed that FOXO4 is elevated in senescent cells and maintains their viability by preventing p<sup>53</sup>-induced apoptosis. Inhibition of the interaction between FOXO4 and p<sup>53</sup> by a modified peptide (FOXO4-DRI [D-retro-inverso]) resulted in p<sup>53</sup> induced apoptosis of senescent cells but also improved fitness, fur density and renal function in both naturally aged mice, as well as in a premature aging (Xpd<sup>TTD/TTD</sup>) model (4). Whether substances like FOXO4-DRI could also have an impact on cellular senescence in the uremic phenotype should be investigated in future studies.

A special type of cellular senescence, which may contribute to uremic inflammation, is *immunosenescence of the adaptive immune system*. An increase in pro-inflammatory CD4<sup>+</sup> CD28<sup>-</sup> effector cells and an imbalance of the T<sub>reg</sub>/TH17 cell ratio, simulating immunosenescence, has been detected in uremic serum (8, 26, 173). CD4<sup>+</sup> CD25<sup>+</sup> FoxP3<sup>+</sup> T<sub>reg</sub> cells have an inhibiting effect on systemic inflammation by releasing anti-inflammatory cytokines, such as IL-10 and TGF- $\beta$ . Since a relation was observed between CRP and IL-6 with and TH17 frequency and an inverse relation was observed between these factors and T<sub>reg</sub> frequency, a role for T<sub>reg</sub>/TH17 dysregulation in the pathogenesis of uremic inflammation could be suggested (173). It has also been reported that p-cresyl sulfate induces macrophage activation and interfere in antigen processing, which lead to a failure in the uremic adaptive immune response (3). A consequence of *vascular cellular senescence*,

which could potentially be of major relevance in uremia, is breakdown of the blood brain barrier (168). Potentially, this could contribute to passage of retained cytokines and uremic toxins from the circulation to the brain and promote cognitive dysfunction, anorexia, and depression; all common features of the uremic phenotype (36).

#### *Abnormalities in tissue homeostasis*

Abnormalities in tissue homeostasis can also contribute to uremic inflammation following the concept of “para-inflammation” (95). One important potential trigger of uremic inflammation resides in visceral *adipose tissue*. Many ESRD patients show characteristics of “obese sarcopenia”; i.e. a progressive increase in fat mass and a decline in lean tissue mass commonly associated with inflammation (50, 85). The relative increase in (visceral) fat mass may contribute to uremic inflammation (2) via pro-inflammatory adipokines, like leptin and visfatin. However, a recent observational study has actually paradoxically shown a protective effect for higher BMI levels in inflamed, but not in non-inflamed dialysis patients, showing the complexity and reverse causation of pathophysiologic relations that are operative in wasted and inflamed ESRD patients (133).

Abnormalities in fluid or sodium composition of the extracellular tissue could also contribute to uremic inflammation. A relation between extracellular fluid overload and inflammation, as evidenced by CRP or IL-6 levels has been observed in various studies in both HD- and PD-patients (30, 37, 52, 66). Moreover, in accordance with the theory of catalytic effects of inflammation (18) the combined presence of fluid overload and inflammation was associated with a multiplicative risk of mortality as compared to the presence of fluid overload or inflammation in isolation (29). The mechanisms behind the relation between fluid overload and inflammation can theoretically be explained either by increased translocation of endotoxins or gut microbes or microbial fragments across an oedematous bowel wall (i.e. *leaky gut*), or by a progressive decline in lean tissue mass due to sustained inflammation, or by translocation of fluid from the vascular to the interstitial compartments, which may hamper removal during dialysis (29, 55).

349 There is an accumulation of osmotically interchangeable sodium not only in dialysis patients, but also  
350 in patients with non-uremic ageing or uncontrolled hypertension (151). The sodium concentration in  
351 this compartment has been estimated to be around 40 mmol/l greater than measured in plasma (10).  
352 Accumulation of interstitial sodium may act pro-inflammatory by stimulation of monocytes, and  
353 induction of IL-17-producing CD4+ T helper (Th17) cells, which may lead to systemic inflammation  
354 (64). In addition, sodium chloride inhibited the activation of IL-4 and IL-13 stimulating M2 (anti-  
355 inflammatory) macrophages (10), as well the suppressive function of FOXP3+ regulatory T cells (46).  
356 The antibacterial effects of sodium and may be an evolutionary conserved mechanism for  
357 antimicrobial skin defense (53). However, whether interstitial sodium accumulation contributes to  
358 persistent inflammation and/or has a causal role in the pathogenesis of premature ageing in CKD has  
359 not yet been definitely established. A last putative factor contributing to uremic inflammation is  
360 *tissue hypoxia*. Studies in healthy subjects have shown activation of the innate immune system, as  
361 reflected by an increase in IL-6 and CRP, as well as by an increase in natural killer cells in response to  
362 hypoxia (43, 65). Recent evidence indicates that HD-patients suffering from prolonged intradialytic  
363 hypoxemia, a condition defined as arterial oxygen saturation levels  $\leq 90\%$  for more than 1/3 of the  
364 treatment time, exhibit a pro-inflammatory phenotype (98). Low arterial oxygen saturation, anemia,  
365 and low cardiac output are frequently concurrently present in HD patients and may put tissues at an  
366 increased risk for hypoxia. Hypoxia triggers adaptive processes in all nucleated cells. HIF-1 mediates  
367 the expression of glycolytic enzymes and a switch from oxidative to glycolytic metabolism. This  
368 metabolic change results in an increased formation of superoxide, hydrogen peroxide and other toxic  
369 ROS (35, 124). HIF regulates an array of processes associated with the immune response and the host  
370 response to infection; in particular HIF plays a key role in the activities of T cells, B cells, dendritic  
371 cells, macrophages, and neutrophils. Members of the NF $\kappa$ B family regulate inflammation and interact  
372 with members of the PHD (prolyl hydroxylase domain)–HIF pathway in ways that link inflammation to  
373 hypoxia (121). Taken together, given the pro-inflammatory effects of local and systemic hypoxia and  
374 given the novel data there is a distinct possibility that hypoxemia may play a role in the genesis of the



pro-inflammatory uremic phenotype. Emerging data link tissue hypoxia to both mitochondrial dysfunction and inflamed uremic fat (134). Since data from a rodent model of programmed cardiovascular dysfunction link hypoxic pregnancy and oxidative stress to endothelial dysfunction, inflammation and premature aging (1) more research is needed in this area. Lastly, *systemic factors* such as depression, as well as socioeconomic and psychosocial factors and associated epigenetic preconditioning, may contribute to uremic inflammation (92), although it is not yet exactly clear to which extent [Figure 1].

### **Systemic effects of uremic inflammation**

Systemic low-grade inflammation is considered to be a cause of premature aging not only in CKD, but also in other chronic diseases such as chronic obstructive pulmonary disease (COPD), congestive heart failure (CHF), rheumatoid arthritis (RA) and HIV (6, 104, 115, 154). In the previous paragraphs, we have explored to which extent chronic inflammation resembles “inflammaging” in the general population. We outlined how inflammation can contribute to cellular damage as well as activation of cellular stress resistance mechanisms. This can lead to effects in various organ system by a variety of changes, such as endothelial dysfunction, vascular calcification, increased vascular stiffness, left ventricular diastolic dysfunction, osteoporosis, cognitive dysfunction and muscular atrophy (68, 137, 149).

The long-term cumulative effects could lead to various clinical syndromes. The most well known are the malnutrition, inflammation and atherosclerosis (MIA) syndrome (17), and the frailty syndrome [Figure 3], defined by loss of lean tissue mass and muscle weakness as well as a reduced functional capacity, (54). Although the relation between biomarkers of aging and these different phenotypes has not been assessed yet, it is likely that these can be considered a subset of a premature ageing syndrome. Systemic inflammation could also impair the homeostatic balance of the body in various

ways. The first is an impaired functioning and reduced structural reserve of cells and vital tissues by direct damage, by mitochondrial dysfunction, or by relocation of free energy for cellular maintenance and repair to the immune system (142, 146). Secondly, systemic inflammation could also impair homeostasis by influencing regulatory networks of the body. Homeostasis depends on a smooth information transfer at all levels; from individual cells to supersystems (163). Systemic inflammation may impair the normal homeostatic fine regulation through the communication after extracellular spillover of cytokines as well as by an abnormal sympathovagal balance (38) (152), thus prioritizing the inflammatory response over normal homeostatic regulation, as well as inducing allostatic load.

Concluding, systemic inflammation can lead to a reduced structural and functional reserve as well as impaired regulatory mechanisms, resulting in reduced resilience to internal and external stressors. This occurs in combination with the loss of kidney function, which cannot be fully replaced by contemporary dialysis techniques, and with comorbidities. Together, this could provide an explanation for the greatly increased risk of morbidity and mortality in ESRD (67) and, metaphorically stated, for an acceleration of biological time (69, 169).

## **Outlook**

Whereas persistent systemic inflammation appears to be a major contributor to adverse outcomes as well as progeria in ESRD, it is not an inevitable consequence of reduced renal function. Indeed, a significant proportion of patients with ESRD have either normal or varying levels of inflammatory markers (97). Next to further investigations of reversible causes of uremic inflammation, it is also of relevance to try to identify patients that are protected from inflammation. Following the example of respiratory medicine (48), “endotyping” of CKD patients, by which detailed phenotypes are coupled to (epi)genetic variants and other biomarkers, could shed more light on both pro-inflammatory as well as protective mechanisms, and their relation to a premature ageing phenotype. There is a great

opportunity for collaboration between basic as well as clinical researchers on this topic, because, as also shown in this review, the study of uremic inflammation is relevant at all system levels in the body, from (epi)genetics to phenotype. This provides indeed the opportunity to connect these different system levels in order to identify central mechanisms which are ideally also amendable for therapeutic interventions.

However, given the multidimensional causality of uremic inflammation, it is unlikely that a single therapeutic “magic bullet” will ever be identified. Recent reviews (81, 136) summarized five therapeutical concepts, which could be applied to combat inflammation in ESRD. The first is to identify and treat underlying sources of inflammation. The second is to promote healthy dietary habits and lifestyle changes that include low-intensity exercise programs (84). Third, in an experimental setting, pharmacological interventions developed to combat inflammation in other chronic diseases as well as, fourth, anti-cytokine treatments may also be applied in ESRD considered not at increased risk for infectious complications. Finally, as recent data imply that elimination of p<sup>16INK4a</sup> positive cells improve health span in mice (5), senolytic drugs, such as dasatinab and quercetin, should be tested in conditions in which senescence may contribute to disease pathogenesis, such as ESRD. Next to interventions specifically focusing on inflammation, it is also of major importance to increase the resilience of the body by physical activity and adequate diet, next to reducing end organ damage due to allostatic overload by factors other than inflammation, e.g. by adequate fluid and blood pressure control and adequate dialysis technique.

Despite the huge potential of the approaches mentioned above (27), few controlled studies have proven success in the management of oxidative stress or systemic inflammation in the uremic milieu. It is of important to realize that targeted mechanisms may have pleiotropic effects (62), or that targeted interventions might focus on pathways, which are influenced by multiple other factors (41). In the future, it is likely that these pitfalls can be partly avoided by the further elucidation of pro- and anti-inflammatory pathways. Since inflammatory biomarkers are “moving targets”, randomized

controlled trials need to include large number of patients in each arm in order to provide sufficient power to prove any anti-inflammatory effect of various interventions. Moreover, given the strong interaction between inflammation and the progeric process (*"inflammaging"*) it is likely that interventions developed in the gerontology field, or in other chronic diseases will also have relevance for CKD and vice versa (126).

## **Conclusion**

Important conceptual similarities exist between uremic inflammation and *"inflammaging"* in the general population. The native inflammatory system is based on a highly evolutionary preserved mechanism, which shows a common effector response to a variety of noxious stimuli. In this sense, it also shares important similarities with other chronic diseases, such as COPD, CHF, RA and HIV, although there are clearly disease specific phenotypical differences next to important similarities. It might therefore be hypothesized that uremic inflammation is an example of progressive *"unhealthy"* aging, both mechanistically as well as phenotypically. Thus, it could be speculated that age-related diseases could be treated more effectively by modulating fundamental mechanisms of aging *per se*, versus the attempt to prevent or delay organ-specific complications one at a time. Studies incorporating patients with different chronic diseases as well as aging subjects may shed more role in the relation between phenotypes and their underlying mechanisms and could provide an answer the question whether phenotypical alterations in these diseases are indeed an example of progressive unhealthy aging. This could in turn lead to shared and better treatment approaches for *"inflammaging"*.

## **Acknowledgements**

Peter Stenvinkels research benefited by support from Swedish Medical Research Council (VR), Swedish Heart and Lung Foundation (20160384), Njurfonden and Westmans Foundation. Peter Kotanko holds stock options in Fresenius Medical Care.

473

474

## 475 References

- 476 1. **Allison BJ, Kaandorp JJ, Kane AD, Camm EJ, Lusby C, Cross CM, Nevin-Dolan R, Thakor AS,**  
 477 **Derks JB, Tarry-Adkins JL, Ozanne SE, and Giussani DA.** Divergence of mechanistic pathways  
 478 mediating cardiovascular aging and developmental programming of cardiovascular disease. *FASEB J*,  
 479 2016.
- 480 2. **Axelsson J, Heimbürger O, Lindholm B, and Stenvinkel P.** Adipose tissue and its relation to  
 481 inflammation: the role of adipokines. *J Ren Nutr* 15: 131-136, 2005.
- 482 3. **Azevedo ML, Bonan NB, Dias G, Brehm F, Steiner TM, Souza WM, Stinghen AE, Barreto FC,**  
 483 **Elifio-Espósito S, Pecoits-Filho R, and Moreno-Amaral AN.** p-Cresyl sulfate affects the oxidative  
 484 burst, phagocytosis process, and antigen presentation of monocyte-derived macrophages. *Toxicol*  
 485 *Lett* 263: 1-5, 2016.
- 486 4. **Baar MP, Brandt RM, Putavet DA, Klein JD, Derks KW, Bourgeois BR, Stryeck S, Rijksen Y,**  
 487 **van Willigenburg H, Feijtel DA, van der Pluijm I, Essers J, van Cappellen WA, van IWF, Houtsmuller**  
 488 **AB, Pothof J, de Bruin RW, Madl T, Hoeijmakers JH, Campisi J, and de Keizer PL.** Targeted Apoptosis  
 489 of Senescent Cells Restores Tissue Homeostasis in Response to Chemotoxicity and Aging. *Cell* 169:  
 490 132-147 e116, 2017.
- 491 5. **Baker DJ, Childs BG, Durik M, Wijers ME, Sieben CJ, Zhong J, Saltness RA, Jeganathan KB,**  
 492 **Verzosa GC, Pezeshki A, Khazaie K, Miller JD, and van Deursen JM.** Naturally occurring p16(Ink4a)-  
 493 positive cells shorten healthy lifespan. *Nature* 530: 184-189, 2016.
- 494 6. **Barnes PJ.** Senescence in COPD and its Comorbidities. *Annu Rev Physiol*, 2016.
- 495 7. **Bessueille L and Magne D.** Inflammation: a culprit for vascular calcification in atherosclerosis  
 496 and diabetes. *Cell Mol Life Sci* 72: 2475-2489, 2015.
- 497 8. **Betjes MG, Langerak AW, van der Spek A, de Wit EA, and Litjens NH.** Premature aging of  
 498 circulating T cells in patients with end-stage renal disease. *Kidney Int* 80: 208-217, 2011.
- 499 9. **Betjes MG, Meijers RW, and Litjens NH.** Loss of renal function causes premature aging of the  
 500 immune system. *Blood Purif* 36: 173-178, 2013.
- 501 10. **Binger KJ, Gebhardt M, Heinig M, Rintisch C, Schroeder A, Neuhofer W, Hilgers K, Manzel A,**  
 502 **Schwartz C, Kleinewietfeld M, Voelkl J, Schatz V, Linker RA, Lang F, Voehringer D, Wright MD,**  
 503 **Hubner N, Dechend R, Jantsch J, Titze J, and Muller DN.** High salt reduces the activation of IL-4- and  
 504 IL-13-stimulated macrophages. *J Clin Invest* 125: 4223-4238, 2015.
- 505 11. **Bro S, Flyvbjerg A, Binder CJ, Bang CA, Denner L, Olgaard K, and Nielsen LB.** A neutralizing  
 506 antibody against receptor for advanced glycation end products (RAGE) reduces atherosclerosis in  
 507 uremic mice. *Atherosclerosis* 201: 274-280, 2008.
- 508 12. **Bruchfeld A, Qureshi AR, Lindholm B, Barany P, Yang L, Stenvinkel P, and Tracey KJ.** High  
 509 Mobility Group Box Protein-1 correlates with renal function in chronic kidney disease (CKD). *Mol Med*  
 510 14: 109-115, 2008.
- 511 13. **Buendia P, Carracedo J, Soriano S, Madueno JA, Ortiz A, Martin-Malo A, Aljama P, and**  
 512 **Ramirez R.** Klotho Prevents NFκB Translocation and Protects Endothelial Cell From Senescence  
 513 Induced by Uremia. *J Gerontol A Biol Sci Med Sci* 70: 1198-1209, 2015.
- 514 14. **Caballo C, Palomo M, Cases A, Galan AM, Molina P, Vera M, Bosch X, Escolar G, and Diaz-**  
 515 **Ricart M.** NFκB in the development of endothelial activation and damage in uremia: an in vitro  
 516 approach. *PLoS One* 7: e43374, 2012.
- 517 15. **Caglar K, Yilmaz MI, Saglam M, Cakir E, Acikel C, Eyileten T, Yenicesu M, Oguz Y, Vural A,**  
 518 **Carrero JJ, Axelsson J, Lindholm B, and Stenvinkel P.** Short-term treatment with sevelamer increases

serum fetuin-a concentration and improves endothelial dysfunction in chronic kidney disease stage 4 patients. *Clin J Am Soc Nephrol* 3: 61-68, 2008.

16. **Carracedo J, Buendia P, Merino A, Soriano S, Esquivias E, Martin-Malo A, Aljama P, and Ramirez R.** Cellular senescence determines endothelial cell damage induced by uremia. *Exp Gerontol* 48: 766-773, 2013.

17. **Carrero JJ and Stenvinkel P.** Inflammation in end-stage renal disease--what have we learned in 10 years? *Semin Dial* 23: 498-509, 2010.

18. **Carrero JJ and Stenvinkel P.** Persistent inflammation as a catalyst for other risk factors in chronic kidney disease: a hypothesis proposal. *Clin J Am Soc Nephrol* 4 Suppl 1: S49-55, 2009.

19. **Carrero JJ, Stenvinkel P, Fellstrom B, Qureshi AR, Lamb K, Heimbürger O, Barany P, Radhakrishnan K, Lindholm B, Soveri I, Nordfors L, and Shiels PG.** Telomere attrition is associated with inflammation, low fetuin-A levels and high mortality in prevalent haemodialysis patients. *J Intern Med* 263: 302-312, 2008.

20. **Castellino P, Bia M, and DeFronzo RA.** Metabolic response to exercise in dialysis patients. *Kidney Int* 32: 877-883, 1987.

21. **Chen WT, Hung KC, Wen MS, Hsu PY, Chen TH, Wang HD, Fang JT, Shie SS, and Wang CY.** Impaired leukocytes autophagy in chronic kidney disease patients. *Cardiorenal Med* 3: 254-264, 2013.

22. **Chen Z, Qureshi AR, Ripsweden J, Wennberg L, Heimbürger O, Lindholm B, Barany P, Haarhaus M, Brismar TB, and Stenvinkel P.** Vertebral bone density associates with coronary artery calcification and is an independent predictor of poor outcome in end-stage renal disease patients. *Bone* 92: 50-57, 2016.

23. **Childs BG, Durik M, Baker DJ, and van Deursen JM.** Cellular senescence in aging and age-related disease: from mechanisms to therapy. *Nat Med* 21: 1424-1435, 2015.

24. **Costello-White R, Ryff CD, and Coe CL.** Aging and low-grade inflammation reduce renal function in middle-aged and older adults in Japan and the USA. *Age (Dordr)* 37: 9808, 2015.

25. **Dai XY, Zhao MM, Cai Y, Guan QC, Zhao Y, Guan Y, Kong W, Zhu WG, Xu MJ, and Wang X.** Phosphate-induced autophagy counteracts vascular calcification by reducing matrix vesicle release. *Kidney Int* 83: 1042-1051, 2013.

26. **Danyan C, Xiaolong H, Song L, Hua G, Weixue T, and Ke L.** The effects of rhBMP-2 and Treg/Th17 functional disequilibrium in uremic patients with cardiovascular complication after maintenance hemodialysis. *Int J Artif Organs* 36: 464-472, 2013.

27. **de Zeeuw D, Akizawa T, Audhya P, Bakris GL, Chin M, Christ-Schmidt H, Goldsberry A, Houser M, Krauth M, Lambers Heerspink HJ, McMurray JJ, Meyer CJ, Parving HH, Remuzzi G, Toto RD, Vaziri ND, Wanner C, Wittes J, Wroldstad D, Chertow GM, and Investigators BT.** Bardoxolone methyl in type 2 diabetes and stage 4 chronic kidney disease. *N Engl J Med* 369: 2492-2503, 2013.

28. **Deban L, Bottazzi B, Garlanda C, de la Torre YM, and Mantovani A.** Pentraxins: multifunctional proteins at the interface of innate immunity and inflammation. *Biofactors* 35: 138-145, 2009.

29. **Dekker M.** Unravelling the relationship between mortality, hyponatremia, inflammation and malnutrition in hemodialysis patients: results from the international MONDO initiative. *Eur J Clin Nutr* In press, 2016.

30. **Demirci MS, Demirci C, Ozdogan O, Kircelli F, Akcicek F, Basci A, Ok E, and Ozkahya M.** Relations between malnutrition-inflammation-atherosclerosis and volume status. The usefulness of bioimpedance analysis in peritoneal dialysis patients. *Nephrol Dial Transplant* 26: 1708-1716, 2011.

31. **Dickhout JG, Carlisle RE, and Austin RC.** Interrelationship between cardiac hypertrophy, heart failure, and chronic kidney disease: endoplasmic reticulum stress as a mediator of pathogenesis. *Circ Res* 108: 629-642, 2011.

32. **Drew DA, Bhadelia R, Tighiouart H, Novak V, Scott TM, Lou KV, Shaffi K, Weiner DE, and Sarnak MJ.** Anatomic brain disease in hemodialysis patients: a cross-sectional study. *Am J Kidney Dis* 61: 271-278, 2013.

33. **Dungey M, Hull KL, Smith AC, Burton JO, and Bishop NC.** Inflammatory factors and exercise in chronic kidney disease. *Int J Endocrinol* 2013: 569831, 2013.
34. **Ekdahl KN, Soveri I, Hilborn J, Fellstrom B, and Nilsson B.** Cardiovascular disease in haemodialysis: role of the intravascular innate immune system. *Nat Rev Nephrol* 13: 285-296, 2017.
35. **Eltzschig HK and Carmeliet P.** Hypoxia and inflammation. *N Engl J Med* 364: 656-665, 2011.
36. **Erdo F, Denes L, and de Lange E.** Age-associated physiological and pathological changes at the blood-brain barrier: A review. *J Cereb Blood Flow Metab* 37: 4-24, 2017.
37. **Fan S and Davenport A.** Does increased glucose exposure lead to increased body fat and reduced lean body mass in anuric peritoneal dialysis patients? *Eur J Clin Nutr* 68: 1253-1254, 2014.
38. **Franceschi C, Garagnani P, Vitale G, Capri M, and Salvioli S.** Inflammaging and 'Garb-aging'. *Trends Endocrinol Metab*, 2016.
39. **Frost RA and Lang CH.** mTor signaling in skeletal muscle during sepsis and inflammation: where does it all go wrong? *Physiology (Bethesda)* 26: 83-96, 2011.
40. **Galle J, Seibold S, and Wanner C.** Inflammation in uremic patients: what is the link? *Kidney Blood Press Res* 26: 65-75, 2003.
41. **Gardner SE, Humphry M, Bennett MR, and Clarke MC.** Senescent Vascular Smooth Muscle Cells Drive Inflammation Through an Interleukin-1alpha-Dependent Senescence-Associated Secretory Phenotype. *Arterioscler Thromb Vasc Biol* 35: 1963-1974, 2015.
42. **Granata S, Masola V, Zoratti E, Scupoli MT, Baruzzi A, Messa M, Sallustio F, Gesualdo L, Lupo A, and Zaza G.** NLRP3 inflammasome activation in dialyzed chronic kidney disease patients. *PLoS One* 10: e0122272, 2015.
43. **Hartmann G, Tschop M, Fischer R, Bidlingmaier C, Riepl R, Tschop K, Hautmann H, Endres S, and Toepfer M.** High altitude increases circulating interleukin-6, interleukin-1 receptor antagonist and C-reactive protein. *Cytokine* 12: 246-252, 2000.
44. **Hellberg M, Hoglund P, Svensson P, Abdulahi H, and Clyne N.** A decline in measured GFR is associated with a decrease in endurance, strength, balance and fine motor skills. *Nephrology (Carlton)*, 2016.
45. **Hermans MM, Brandenburg V, Ketteler M, Kooman JP, van der Sande FM, Boeschoten EW, Leunissen KM, Krediet RT, Dekker FW, and Netherlands cooperative study on the adequacy of D.** Association of serum fetuin-A levels with mortality in dialysis patients. *Kidney Int* 72: 202-207, 2007.
46. **Hernandez AL, Kitz A, Wu C, Lowther DE, Rodriguez DM, Vudattu N, Deng S, Herold KC, Kuchroo VK, Kleinewietfeld M, and Hafler DA.** Sodium chloride inhibits the suppressive function of FOXP3+ regulatory T cells. *J Clin Invest* 125: 4212-4222, 2015.
47. **Himmelfarb J, Stenvinkel P, Ikizler TA, and Hakim RM.** The elephant in uremia: oxidant stress as a unifying concept of cardiovascular disease in uremia. *Kidney Int* 62: 1524-1538, 2002.
48. **Hizawa N.** Clinical approaches towards asthma and chronic obstructive pulmonary disease based on the heterogeneity of disease pathogenesis. *Clin Exp Allergy* 46: 678-687, 2016.
49. **Holmen C, Elsheikh E, Stenvinkel P, Qureshi AR, Pettersson E, Jalkanen S, and Sumitran-Holgersson S.** Circulating inflammatory endothelial cells contribute to endothelial progenitor cell dysfunction in patients with vasculitis and kidney involvement. *J Am Soc Nephrol* 16: 3110-3120, 2005.
50. **Honda H, Qureshi AR, Axelsson J, Heimbürger O, Suliman ME, Barany P, Stenvinkel P, and Lindholm B.** Obese sarcopenia in patients with end-stage renal disease is associated with inflammation and increased mortality. *Am J Clin Nutr* 86: 633-638, 2007.
51. **Izquierdo MC, Perez-Gomez MV, Sanchez-Nino MD, Sanz AB, Ruiz-Andres O, Poveda J, Moreno JA, Egido J, and Ortiz A.** Klotho, phosphate and inflammation/ageing in chronic kidney disease. *Nephrol Dial Transplant* 27 Suppl 4: iv6-10, 2012.
52. **Jacobs LH, van de Kerkhof JJ, Mingels AM, Passos VL, Kleijnen VW, Mazairac AH, van der Sande FM, Wodzig WK, Konings CJ, Leunissen KM, van Dieijen-Visser MP, and Kooman JP.** Inflammation, overhydration and cardiac biomarkers in haemodialysis patients: a longitudinal study. *Nephrol Dial Transplant* 25: 243-248, 2010.

53. **Jantsch J, Schatz V, Friedrich D, Schroder A, Kopp C, Siegert I, Maronna A, Wendelborn D, Linz P, Binger KJ, Gebhardt M, Heinig M, Neubert P, Fischer F, Teufel S, David JP, Neufert C, Cavallaro A, Rakova N, Kuper C, Beck FX, Neuhofer W, Muller DN, Schuler G, Uder M, Bogdan C, Luft FC, and Titze J.** Cutaneous Na<sup>+</sup> storage strengthens the antimicrobial barrier function of the skin and boosts macrophage-driven host defense. *Cell Metab* 21: 493-501, 2015.
54. **Johansen KL, Chertow GM, Jin C, and Kutner NG.** Significance of frailty among dialysis patients. *J Am Soc Nephrol* 18: 2960-2967, 2007.
55. **John B, Tan BK, Dainty S, Spanel P, Smith D, and Davies SJ.** Plasma volume, albumin, and fluid status in peritoneal dialysis patients. *Clin J Am Soc Nephrol* 5: 1463-1470, 2010.
56. **Jurk D, Wilson C, Passos JF, Oakley F, Correia-Melo C, Greaves L, Saretzki G, Fox C, Lawless C, Anderson R, Hewitt G, Pender SL, Fullard N, Nelson G, Mann J, van de Sluis B, Mann DA, and von Zglinicki T.** Chronic inflammation induces telomere dysfunction and accelerates ageing in mice. *Nat Commun* 2: 4172, 2014.
57. **Kanda N, Matsui K, Kawai T, Edamatsu H, Tanuma Y, Suzuki O, Takahashi T, and Kamakura S.** Implantation of octacalcium phosphate collagen composites (OCP/Col) after extraction of canine deciduous teeth achieved undisturbed permanent tooth eruption. *Arch Oral Biol* 72: 179-186, 2016.
58. **Kato S, Chmielewski M, Honda H, Pecoits-Filho R, Matsuo S, Yuzawa Y, Traaneus A, Stenvinkel P, and Lindholm B.** Aspects of immune dysfunction in end-stage renal disease. *Clin J Am Soc Nephrol* 3: 1526-1533, 2008.
59. **Kauppinen A, Suuronen T, Ojala J, Kaarniranta K, and Salminen A.** Antagonistic crosstalk between NF-kappaB and SIRT1 in the regulation of inflammation and metabolic disorders. *Cell Signal* 25: 1939-1948, 2013.
60. **Kaysen GA.** Inflammation and oxidative stress in end-stage renal disease. *Adv Nephrol Necker Hosp* 30: 201-214, 2000.
61. **Keyzer CA, de Borst MH, van den Berg E, Jahnen-Dechent W, Arampatzis S, Farese S, Bergmann IP, Floege J, Navis G, Bakker SJ, van Goor H, Eisenberger U, and Pasch A.** Calcification Propensity and Survival among Renal Transplant Recipients. *J Am Soc Nephrol* 27: 239-248, 2016.
62. **Kim HJ and Vaziri ND.** Contribution of impaired Nrf2-Keap1 pathway to oxidative stress and inflammation in chronic renal failure. *Am J Physiol Renal Physiol* 298: F662-671, 2010.
63. **Kirk JE.** The glyoxalase I activity of arterial tissue in individuals of various ages. *J Gerontol* 15: 139-141, 1960.
64. **Kleinewietfeld M, Manzel A, Titze J, Kvakan H, Yosef N, Linker RA, Muller DN, and Hafler DA.** Sodium chloride drives autoimmune disease by the induction of pathogenic TH17 cells. *Nature* 496: 518-522, 2013.
65. **Klokke M, Kharazmi A, Galbo H, Bygbjerg I, and Pedersen BK.** Influence of in vivo hypobaric hypoxia on function of lymphocytes, neutrocytes, natural killer cells, and cytokines. *J Appl Physiol* (1985) 74: 1100-1106, 1993.
66. **Konings CJ, Kooman JP, Schonck M, Struijk DG, Gladziwa U, Hoorntje SJ, van der Wall Bake AW, van der Sande FM, and Leunissen KM.** Fluid status in CAPD patients is related to peritoneal transport and residual renal function: evidence from a longitudinal study. *Nephrol Dial Transplant* 18: 797-803, 2003.
67. **Kooman JP, Broers NJ, Usvyat L, Thijssen S, van der Sande FM, Cornelis T, Levin NW, Leunissen KM, and Kotanko P.** Out of control: accelerated aging in uremia. *Nephrol Dial Transplant* 28: 48-54, 2013.
68. **Kooman JP, Kotanko P, Schols AM, Shiels PG, and Stenvinkel P.** Chronic kidney disease and premature ageing. *Nat Rev Nephrol* 10: 732-742, 2014.
69. **Kooman JP, Usvyat L, van der Sande FM, Thijssen S, Levin N, Leunissen KM, and Kotanko P.** 'Time and time again': oscillatory and longitudinal time patterns in dialysis patients. *Kidney Blood Press Res* 35: 534-548, 2012.
70. **Kramann R, Goettsch C, Wongboonsin J, Iwata H, Schneider RK, Kuppe C, Kaesler N, Chang-Panesso M, Machado FG, Gratwohl S, Madhurima K, Hutcheson JD, Jain S, Aikawa E, and**



**Humphreys BD.** Adventitial MSC-like Cells Are Progenitors of Vascular Smooth Muscle Cells and Drive Vascular Calcification in Chronic Kidney Disease. *Cell Stem Cell* 19: 628-642, 2016.

71. **Kshirsagar AV, Craig RG, Beck JD, Moss K, Offenbacher S, Kotanko P, Yoshino M, Levin NW, Yip JK, Almas K, Lupovici E, and Falk RJ.** Severe periodontitis is associated with low serum albumin among patients on maintenance hemodialysis therapy. *Clin J Am Soc Nephrol* 2: 239-244, 2007.

72. **Kshirsagar AV, Craig RG, Moss KL, Beck JD, Offenbacher S, Kotanko P, Klemmer PJ, Yoshino M, Levin NW, Yip JK, Almas K, Lupovici EM, Usvyat LA, and Falk RJ.** Periodontal disease adversely affects the survival of patients with end-stage renal disease. *Kidney Int* 75: 746-751, 2009.

73. **Kuro-o M.** Klotho, phosphate and FGF-23 in ageing and disturbed mineral metabolism. *Nat Rev Nephrol* 9: 650-660, 2013.

74. **Lau WL, Liu SM, Pahlevan S, Yuan J, Khazaeli M, Ni Z, Chan JY, and Vaziri ND.** Role of Nrf2 dysfunction in uremia-associated intestinal inflammation and epithelial barrier disruption. *Dig Dis Sci* 60: 1215-1222, 2015.

75. **Lee CT, Tsai YC, Ng HY, Su Y, Lee WC, Lee LC, Chiou TT, Liao SC, and Hsu KT.** Association between C-reactive protein and biomarkers of bone and mineral metabolism in chronic hemodialysis patients: a cross-sectional study. *J Ren Nutr* 19: 220-227, 2009.

76. **Leemans JC, Kors L, Anders HJ, and Florquin S.** Pattern recognition receptors and the inflammasome in kidney disease. *Nat Rev Nephrol* 10: 398-414, 2014.

77. **Li G, Tang D, and Lotze MT.** Menage a Trois in stress: DAMPs, redox and autophagy. *Semin Cancer Biol* 23: 380-390, 2013.

78. **Libetta C, Sepe V, Esposito P, Galli F, and Dal Canton A.** Oxidative stress and inflammation: Implications in uremia and hemodialysis. *Clin Biochem* 44: 1189-1198, 2011.

79. **Liu J, Huang K, Cai GY, Chen XM, Yang JR, Lin LR, Yang J, Huo BG, Zhan J, and He YN.** Receptor for advanced glycation end-products promotes premature senescence of proximal tubular epithelial cells via activation of endoplasmic reticulum stress-dependent p21 signaling. *Cell Signal* 26: 110-121, 2014.

80. **Lopez-Otin C, Blasco MA, Partridge L, Serrano M, and Kroemer G.** The hallmarks of aging. *Cell* 153: 1194-1217, 2013.

81. **Machowska A, Carrero JJ, Lindholm B, and Stenvinkel P.** Therapeutics targeting persistent inflammation in chronic kidney disease. *Transl Res* 167: 204-213, 2016.

82. **Maessen DE, Stehouwer CD, and Schalkwijk CG.** The role of methylglyoxal and the glyoxalase system in diabetes and other age-related diseases. *Clin Sci (Lond)* 128: 839-861, 2015.

83. **Mafrà D and Fouque D.** Gut microbiota and inflammation in chronic kidney disease patients. *Clin Kidney J* 8: 332-334, 2015.

84. **Manfredini F, Mallamaci F, D'Arrigo G, Baggetta R, Bolignano D, Torino C, Lamberti N, Bertoli S, Ciurlino D, Rocca-Rey L, Barilla A, Battaglia Y, Rapana RM, Zuccala A, Bonanno G, Fatuzzo P, Rapisarda F, Rastelli S, Fabrizi F, Messa P, De Paola L, Lombardi L, Cupisti A, Fuiano G, Lucisano G, Summaria C, Felisatti M, Pozzato E, Malagoni AM, Castellino P, Aucella F, ElHafeez SA, Provenzano PF, Tripepi G, Catizone L, and Zoccali C.** Exercise in Patients on Dialysis: A Multicenter, Randomized Clinical Trial. *J Am Soc Nephrol* 28: 1259-1268, 2017.

85. **Marcelli D, Brand K, Ponce P, Milkowski A, Marelli C, Ok E, Merello Godino JI, Gurevich K, Jirka T, Rosenberger J, Di Benedetto A, Ladanyi E, Grassmann A, Scatizzi L, Bayh I, Kooman J, and Canaud B.** Longitudinal Changes in Body Composition in Patients After Initiation of Hemodialysis Therapy: Results From an International Cohort. *J Ren Nutr*, 2015.

86. **Marcelli D, Usvyat LA, Kotanko P, Bayh I, Canaud B, Etter M, Gatti E, Grassmann A, Wang Y, Marelli C, Scatizzi L, Stopper A, van der Sande FM, Kooman J, and Consortium MODO.** Body composition and survival in dialysis patients: results from an international cohort study. *Clin J Am Soc Nephrol* 10: 1192-1200, 2015.

87. **Margolick JB and Ferrucci L.** Accelerating aging research: how can we measure the rate of biologic aging? *Exp Gerontol* 64: 78-80, 2015.

88. **Martens RJ, Kooman JP, Stehouwer CD, Dagnelie PC, van der Kallen CJ, Koster A, Kroon AA, Leunissen KM, Nijpels G, van der Sande FM, Schaper NC, Sep SJ, van Bortel MP, Schram MT, and**

**Henry RM.** Estimated GFR, Albuminuria, and Cognitive Performance: The Maastricht Study. *Am J Kidney Dis*, 2016.

89. **Martin-Rodriguez S, Caballo C, Gutierrez G, Vera M, Cruzado JM, Cases A, Escolar G, and Diaz-Ricart M.** TLR4 and NALP3 inflammasome in the development of endothelial dysfunction in uraemia. *Eur J Clin Invest* 45: 160-169, 2015.

90. **McCullough PA and Ali S.** Cardiac and renal function in patients with type 2 diabetes who have chronic kidney disease: potential effects of bardoxolone methyl. *Drug Des Devel Ther* 6: 141-149, 2012.

91. **McGuinness D, Leierer J, Shapter O, Mohammed S, Gingell-Littlejohn M, Kingsmore DB, Little AM, Kerschbaum J, Schneeberger S, Maglione M, Nadalin S, Wagner S, Konigsrainer A, Aitken E, Whalen H, Clancy M, McConnachie A, Koppelstaetter C, Stevenson KS, and Shiels PG.** Identification of Molecular Markers of Delayed Graft Function Based on the Regulation of Biological Ageing. *PLoS One* 11: e0146378, 2016.

92. **McGuinness D, McGlynn LM, Johnson PC, MacIntyre A, Batty GD, Burns H, Cavanagh J, Deans KA, Ford I, McConnachie A, McGinty A, McLean JS, Millar K, Packard CJ, Sattar NA, Tannahill C, Velupillai YN, and Shiels PG.** Socio-economic status is associated with epigenetic differences in the pSoBid cohort. *Int J Epidemiol* 41: 151-160, 2012.

93. **McIntyre CW, Harrison LE, Eldehni MT, Jefferies HJ, Szeto CC, John SG, Sigrist MK, Burton JO, Hothi D, Korsheed S, Owen PJ, Lai KB, and Li PK.** Circulating endotoxemia: a novel factor in systemic inflammation and cardiovascular disease in chronic kidney disease. *Clin J Am Soc Nephrol* 6: 133-141, 2011.

94. **McLellan AC and Thornalley PJ.** Glyoxalase activity in human red blood cells fractioned by age. *Mech Ageing Dev* 48: 63-71, 1989.

95. **Medzhitov R.** Origin and physiological roles of inflammation. *Nature* 454: 428-435, 2008.

96. **Medzhitov R and Horng T.** Transcriptional control of the inflammatory response. *Nat Rev Immunol* 9: 692-703, 2009.

97. **Meuwese CL, Stenvinkel P, Dekker FW, and Carrero JJ.** Monitoring of inflammation in patients on dialysis: forewarned is forearmed. *Nat Rev Nephrol* 7: 166-176, 2011.

98. **Meyring-Wosten A, Zhang H, Ye X, Fuertinger DH, Chan L, Kappel F, Artemyev M, Ginsberg N, Wang Y, Thijssen S, and Kotanko P.** Intradialytic Hypoxemia and Clinical Outcomes in Patients on Hemodialysis. *Clin J Am Soc Nephrol*, 2016.

99. **Missailidis C, Hallqvist J, Qureshi AR, Barany P, Heimbürger O, Lindholm B, Stenvinkel P, and Bergman P.** Serum Trimethylamine-N-Oxide Is Strongly Related to Renal Function and Predicts Outcome in Chronic Kidney Disease. *PLoS One* 11: e0141738, 2016.

100. **Morcos M, Du X, Pfisterer F, Hutter H, Sayed AA, Thornalley P, Ahmed N, Baynes J, Thorpe S, Kukudov G, Schlotterer A, Bozorgmehr F, El Baki RA, Stern D, Moehrlen F, Ibrahim Y, Oikonomou D, Hamann A, Becker C, Zeier M, Schwenger V, Miftari N, Humpert P, Hammes HP, Buechler M, Bierhaus A, Brownlee M, and Nawroth PP.** Glyoxalase-1 prevents mitochondrial protein modification and enhances lifespan in *Caenorhabditis elegans*. *Aging Cell* 7: 260-269, 2008.

101. **Moreno JA, Izquierdo MC, Sanchez-Nino MD, Suarez-Alvarez B, Lopez-Larrea C, Jakubowski A, Blanco J, Ramirez R, Selgas R, Ruiz-Ortega M, Egido J, Ortiz A, and Sanz AB.** The inflammatory cytokines TWEAK and TNFalpha reduce renal klotho expression through NFkappaB. *J Am Soc Nephrol* 22: 1315-1325, 2011.

102. **Morigi M, Perico L, Rota C, Longaretti L, Conti S, Rottoli D, Novelli R, Remuzzi G, and Benigni A.** Sirtuin 3-dependent mitochondrial dynamic improvements protect against acute kidney injury. *J Clin Invest* 125: 715-726, 2015.

103. **Muteliefu G, Shimizu H, Enomoto A, Nishijima F, Takahashi M, and Niwa T.** Indoxyl sulfate promotes vascular smooth muscle cell senescence with upregulation of p53, p21, and prelamin A through oxidative stress. *Am J Physiol Cell Physiol* 303: C126-134, 2012.

104. **Nasi M, De Biasi S, Gibellini L, Bianchini E, Pecorini S, Bacca V, Guaraldi G, Mussini C, Pinti M, and Cossarizza A.** Ageing and inflammation in patients with HIV infection. *Clin Exp Immunol* 187: 44-52, 2017.

105. **Oh HJ, Nam BY, Lee MJ, Kim CH, Koo HM, Doh FM, Han JH, Kim EJ, Han JS, Park JT, Yoo TH, Kang SW, Han DS, and Han SH.** Decreased circulating klotho levels in patients undergoing dialysis and relationship to oxidative stress and inflammation. *Perit Dial Int* 35: 43-51, 2015.
106. **Ozkok A, Aktas E, Yilmaz A, Telci A, Oflaz H, Deniz G, and Yildiz A.** Decrease in endothelial progenitor cells associated with inflammation, but not with endothelial dysfunction in chronic hemodialysis patients. *Clin Nephrol* 79: 21-30, 2013.
107. **Pedruzzi LM, Cardozo LF, Daleprane JB, Stockler-Pinto MB, Monteiro EB, Leite M, Jr., Vaziri ND, and Mafra D.** Systemic inflammation and oxidative stress in hemodialysis patients are associated with down-regulation of Nrf2. *J Nephrol* 28: 495-501, 2015.
108. **Poggioli S, Bakala H, and Friguet B.** Age-related increase of protein glycation in peripheral blood lymphocytes is restricted to preferential target proteins. *Exp Gerontol* 37: 1207-1215, 2002.
109. **Raj DS, Carrero JJ, Shah VO, Qureshi AR, Barany P, Heimbürger O, Lindholm B, Ferguson J, Moseley PL, and Stenvinkel P.** Soluble CD14 levels, interleukin 6, and mortality among prevalent hemodialysis patients. *Am J Kidney Dis* 54: 1072-1080, 2009.
110. **Ramezani A, Massy ZA, Meijers B, Evenepoel P, Vanholder R, and Raj DS.** Role of the Gut Microbiome in Uremia: A Potential Therapeutic Target. *Am J Kidney Dis*, 2015.
111. **Ramirez R, Carracedo J, Soriano S, Jimenez R, Martin-Malo A, Rodriguez M, Blasco M, and Aljama P.** Stress-induced premature senescence in mononuclear cells from patients on long-term hemodialysis. *Am J Kidney Dis* 45: 353-359, 2005.
112. **Rodier F and Campisi J.** Four faces of cellular senescence. *J Cell Biol* 192: 547-556, 2011.
113. **Rosin DL and Okusa MD.** Dangers within: DAMP responses to damage and cell death in kidney disease. *J Am Soc Nephrol* 22: 416-425, 2011.
114. **Rubio-Ruiz ME, Peredo-Escarcega AE, Cano-Martinez A, and Guarner-Lans V.** An Evolutionary Perspective of Nutrition and Inflammation as Mechanisms of Cardiovascular Disease. *Int J Evol Biol* 2015: 179791, 2015.
115. **Rutten EP, Gopal P, Wouters EF, Franssen FM, Hageman GJ, Vanfleteren LE, Spruit MA, and Reynaert NL.** Various Mechanistic Pathways Representing the Aging Process Are Altered in COPD. *Chest* 149: 53-61, 2016.
116. **Salminen A, Huuskonen J, Ojala J, Kauppinen A, Kaarniranta K, and Suuronen T.** Activation of innate immunity system during aging: NF- $\kappa$ B signaling is the molecular culprit of inflamm-aging. *Ageing Res Rev* 7: 83-105, 2008.
117. **Salminen A and Kaarniranta K.** ER stress and hormetic regulation of the aging process. *Ageing Res Rev* 9: 211-217, 2010.
118. **Salminen A, Kaarniranta K, and Kauppinen A.** Inflammaging: disturbed interplay between autophagy and inflammasomes. *Aging (Albany NY)* 4: 166-175, 2012.
119. **Salminen A, Ojala J, Kaarniranta K, and Kauppinen A.** Mitochondrial dysfunction and oxidative stress activate inflammasomes: impact on the aging process and age-related diseases. *Cell Mol Life Sci* 69: 2999-3013, 2012.
120. **Schaefer L.** Complexity of danger: the diverse nature of damage-associated molecular patterns. *J Biol Chem* 289: 35237-35245, 2014.
121. **Schaffer K and Taylor CT.** The impact of hypoxia on bacterial infection. *FEBS J* 282: 2260-2266, 2015.
122. **Schlotterer A, Kukudov G, Bozorgmehr F, Hutter H, Du X, Oikonomou D, Ibrahim Y, Pfisterer F, Rabbani N, Thornalley P, Sayed A, Fleming T, Humpert P, Schwenger V, Zeier M, Hamann A, Stern D, Brownlee M, Bierhaus A, Nawroth P, and Morcos M.** C. elegans as model for the study of high glucose-mediated life span reduction. *Diabetes* 58: 2450-2456, 2009.
123. **Schmitt R, Susnik N, and Melk A.** Molecular aspects of renal senescence. *Curr Opin Organ Transplant* 20: 412-416, 2015.
124. **Semenza GL.** Oxygen sensing, homeostasis, and disease. *N Engl J Med* 365: 537-547, 2011.
125. **Shi K, Wang F, Jiang H, Liu H, Wei M, Wang Z, and Xie L.** Gut bacterial translocation may aggravate microinflammation in hemodialysis patients. *Dig Dis Sci* 59: 2109-2117, 2014.

126. **Shiels P SPKJAMD.** Circulating markers of ageing and allostatic load: A slow train coming. *Pract Lab Med* 2016.
127. **Shiels PG, McGlynn LM, MacIntyre A, Johnson PC, Batty GD, Burns H, Cavanagh J, Deans KA, Ford I, McConnachie A, McGinty A, McLean JS, Millar K, Sattar N, Tannahill C, Velupillai YN, and Packard CJ.** Accelerated telomere attrition is associated with relative household income, diet and inflammation in the pSoBid cohort. *PLoS One* 6: e22521, 2011.
128. **Smith ER, Cai MM, McMahon LP, Pedagogos E, Toussaint ND, Brumby C, and Holt SG.** Serum fetuin-A concentration and fetuin-A-containing calciprotein particles in patients with chronic inflammatory disease and renal failure. *Nephrology (Carlton)* 18: 215-221, 2013.
129. **Smith ER, Ford ML, Tomlinson LA, Bodenham E, McMahon LP, Farese S, Rajkumar C, Holt SG, and Pasch A.** Serum calcification propensity predicts all-cause mortality in predialysis CKD. *J Am Soc Nephrol* 25: 339-348, 2014.
130. **Smith ER, Hanssen E, McMahon LP, and Holt SG.** Fetuin-A-containing calciprotein particles reduce mineral stress in the macrophage. *PLoS One* 8: e60904, 2013.
131. **Sosa Pena MD, Lopez-Soler R, and Melendez JA.** Senescence in chronic allograft nephropathy. *Am J Physiol Renal Physiol*: ajprenal 00195 02016, 2016.
132. **Stenvinkel P and Alvestrand A.** Inflammation in end-stage renal disease: sources, consequences, and therapy. *Semin Dial* 15: 329-337, 2002.
133. **Stenvinkel P, Gillespie IA, Tunks J, Addison J, Kronenberg F, Drueke TB, Marcelli D, Scherthaner G, Eckardt KU, Floege J, Froissart M, Anker SD, and Committee AROS.** Inflammation Modifies the Paradoxical Association between Body Mass Index and Mortality in Hemodialysis Patients. *J Am Soc Nephrol*, 2016.
134. **Stenvinkel P and Haase VH.** Inflamed fat and mitochondrial dysfunction in end-stage renal disease links to hypoxia-could curcumin be of benefit? *Nephrol Dial Transplant* 32: 909-912, 2017.
135. **Stenvinkel P, Ketteler M, Johnson RJ, Lindholm B, Pecoits-Filho R, Riella M, Heimbürger O, Cederholm T, and Girndt M.** IL-10, IL-6, and TNF-alpha: central factors in the altered cytokine network of uremia--the good, the bad, and the ugly. *Kidney Int* 67: 1216-1233, 2005.
136. **Stenvinkel P, Kooman JP, and Shiels PG.** Nutrients and ageing: what can we learn about ageing interactions from animal biology? *Curr Opin Clin Nutr Metab Care* 19: 19-25, 2016.
137. **Stenvinkel P and Larsson TE.** Chronic kidney disease: a clinical model of premature aging. *Am J Kidney Dis* 62: 339-351, 2013.
138. **Stenvinkel P, Luttropp K, McGuinness D, Witasp A, Qureshi AR, Wernerson A, Nordfors L, Schalling M, Ripsweden J, Wennberg L, Soderberg M, Barany P, Olauson H, and Shiels PG.** CDKN2A/p16INK4a expression is associated with vascular progeria in chronic kidney disease. *Aging (Albany NY)* 9: 494-507, 2017.
139. **Stenvinkel P, Wang K, Qureshi AR, Axelsson J, Pecoits-Filho R, Gao P, Barany P, Lindholm B, Jogestrand T, Heimbürger O, Holmes C, Schalling M, and Nordfors L.** Low fetuin-A levels are associated with cardiovascular death: Impact of variations in the gene encoding fetuin. *Kidney Int* 67: 2383-2392, 2005.
140. **Steyers CM, 3rd and Miller FJ, Jr.** Endothelial dysfunction in chronic inflammatory diseases. *Int J Mol Sci* 15: 11324-11349, 2014.
141. **Stinghen AE, Massy ZA, Vlassara H, Striker GE, and Boullier A.** Uremic Toxicity of Advanced Glycation End Products in CKD. *J Am Soc Nephrol* 27: 354-370, 2016.
142. **Straub RH and Schradin C.** Chronic inflammatory systemic diseases: An evolutionary trade-off between acutely beneficial but chronically harmful programs. *Evol Med Public Health* 2016: 37-51, 2016.
143. **Sturmelechner I, Durik M, Sieben CJ, Baker DJ, and van Deursen JM.** Cellular senescence in renal ageing and disease. *Nat Rev Nephrol* 13: 77-89, 2017.
144. **Sun CY, Chang SC, and Wu MS.** Suppression of Klotho expression by protein-bound uremic toxins is associated with increased DNA methyltransferase expression and DNA hypermethylation. *Kidney Int* 81: 640-650, 2012.

145. **Sun YM, Lin KY, and Chen YQ.** Diverse functions of miR-125 family in different cell contexts. *J Hematol Oncol* 6: 6, 2013.
146. **Supinski GS and Callahan LA.** Free radical-mediated skeletal muscle dysfunction in inflammatory conditions. *J Appl Physiol (1985)* 102: 2056-2063, 2007.
147. **Takata Y, Kitami Y, Yang ZH, Nakamura M, Okura T, and Hiwada K.** Vascular inflammation is negatively autoregulated by interaction between CCAAT/enhancer-binding protein-delta and peroxisome proliferator-activated receptor-gamma. *Circ Res* 91: 427-433, 2002.
148. **Tanaka Y, Joki N, Hase H, Iwasaki M, Ikeda M, Ando R, Shinoda T, Inaguma D, Sakaguchi T, Komatsu Y, Koiwa F, Yamaka T, and Shigematsu T.** Effect of erythropoietin-stimulating agent on uremic inflammation. *J Inflamm (Lond)* 9: 17, 2012.
149. **Ter Maaten JM, Damman K, Verhaar MC, Paulus WJ, Duncker DJ, Cheng C, van Heerebeek L, Hillege HL, Lam CS, Navis G, and Voors AA.** Connecting heart failure with preserved ejection fraction and renal dysfunction: the role of endothelial dysfunction and inflammation. *Eur J Heart Fail* 18: 588-598, 2016.
150. **Thevaranjan N, Puchta A, Schulz C, Naidoo A, Szamosi JC, Verschoor CP, Loukov D, Schenck LP, Jury J, Foley KP, Schertzer JD, Larche MJ, Davidson DJ, Verdu EF, Surette MG, and Bowdish DM.** Age-Associated Microbial Dysbiosis Promotes Intestinal Permeability, Systemic Inflammation, and Macrophage Dysfunction. *Cell Host Microbe* 21: 455-466 e454, 2017.
151. **Titze J, Dahlmann A, Lerchl K, Kopp C, Rakova N, Schroder A, and Luft FC.** Spooky sodium balance. *Kidney Int* 85: 759-767, 2014.
152. **Underwood CF, Hildreth CM, Wyse BF, Boyd R, Goodchild AK, and Phillips JK.** Uraemia: an unrecognized driver of central neurohumoral dysfunction in chronic kidney disease? *Acta Physiol (Oxf)* 219: 305-323, 2017.
153. **van den Ham EC, Kooman JP, Schols AM, Nieman FH, Does JD, Franssen FM, Akkermans MA, Janssen PP, and van Hooff JP.** Similarities in skeletal muscle strength and exercise capacity between renal transplant and hemodialysis patients. *Am J Transplant* 5: 1957-1965, 2005.
154. **van Onna M and Boonen A.** The challenging interplay between rheumatoid arthritis, ageing and comorbidities. *BMC Musculoskelet Disord* 17: 184, 2016.
155. **Vaziri ND, Pahl MV, Crum A, and Norris K.** Effect of uremia on structure and function of immune system. *J Ren Nutr* 22: 149-156, 2012.
156. **Vaziri ND, Wong J, Pahl M, Piceno YM, Yuan J, DeSantis TZ, Ni Z, Nguyen TH, and Andersen GL.** Chronic kidney disease alters intestinal microbial flora. *Kidney Int* 83: 308-315, 2013.
157. **Vaziri ND, Yuan J, Rahimi A, Ni Z, Said H, and Subramanian VS.** Disintegration of colonic epithelial tight junction in uremia: a likely cause of CKD-associated inflammation. *Nephrol Dial Transplant* 27: 2686-2693, 2012.
158. **Wang AY, Sea MM, Tang N, Lam CW, Chan IH, Lui SF, Sanderson JE, and Woo J.** Energy intake and expenditure profile in chronic peritoneal dialysis patients complicated with circulatory congestion. *Am J Clin Nutr* 90: 1179-1184, 2009.
159. **Wang DT, Yang YJ, Huang RH, Zhang ZH, and Lin X.** Myostatin Activates the Ubiquitin-Proteasome and Autophagy-Lysosome Systems Contributing to Muscle Wasting in Chronic Kidney Disease. *Oxid Med Cell Longev* 2015: 684965, 2015.
160. **Watroba M and Szukiewicz D.** The role of sirtuins in aging and age-related diseases. *Adv Med Sci* 61: 52-62, 2016.
161. **White WE, Yaqoob MM, and Harwood SM.** Aging and uremia: Is there cellular and molecular crossover? *World J Nephrol* 4: 19-30, 2015.
162. **Witko-Sarsat V, Friedlander M, Nguyen Khoa T, Capeillere-Blandin C, Nguyen AT, Canteloup S, Dayer JM, Jungers P, Drueke T, and Descamps-Latscha B.** Advanced oxidation protein products as novel mediators of inflammation and monocyte activation in chronic renal failure. *J Immunol* 161: 2524-2532, 1998.
163. **Woods HA and Wilson JK.** An information hypothesis for the evolution of homeostasis. *Trends Ecol Evol* 28: 283-289, 2013.

164. **Xue M, Rabbani N, and Thornalley PJ.** Glyoxalase in ageing. *Semin Cell Dev Biol* 22: 293-301, 2011.
165. **Yamada S, Tatsumoto N, Tokumoto M, Noguchi H, Ooboshi H, Kitazono T, and Tsuruya K.** Phosphate binders prevent phosphate-induced cellular senescence of vascular smooth muscle cells and vascular calcification in a modified, adenine-based uremic rat model. *Calcif Tissue Int* 96: 347-358, 2015.
166. **Yamada S, Tokumoto M, Tatsumoto N, Taniguchi M, Noguchi H, Nakano T, Masutani K, Ooboshi H, Tsuruya K, and Kitazono T.** Phosphate overload directly induces systemic inflammation and malnutrition as well as vascular calcification in uremia. *Am J Physiol Renal Physiol* 306: F1418-1428, 2014.
167. **Yamada S, Tokumoto M, Tsuruya K, Tatsumoto N, Noguchi H, Kitazono T, and Ooboshi H.** Fetuin-A decrease induced by a low-protein diet enhances vascular calcification in uremic rats with hyperphosphatemia. *Am J Physiol Renal Physiol* 309: F744-754, 2015.
168. **Yamazaki Y, Baker DJ, Tachibana M, Liu CC, van Deursen JM, Brott TG, Bu G, and Kanekiyo T.** Vascular Cell Senescence Contributes to Blood-Brain Barrier Breakdown. *Stroke* 47: 1068-1077, 2016.
169. **Yates FE.** Complexity of a human being: changes with age. *Neurobiol Aging* 23: 17-19, 2002.
170. **Zawada AM, Rogacev KS, Muller S, Rotter B, Winter P, Fliser D, and Heine GH.** Massive analysis of cDNA Ends (MACE) and miRNA expression profiling identifies proatherogenic pathways in chronic kidney disease. *Epigenetics* 9: 161-172, 2014.
171. **Zeng W, Guo YH, Qi W, Chen JG, Yang LL, Luo ZF, Mu J, and Feng B.** 4-Phenylbutyric acid suppresses inflammation through regulation of endoplasmic reticulum stress of endothelial cells stimulated by uremic serum. *Life Sci* 103: 15-24, 2014.
172. **Zewinger S, Schumann T, Fliser D, and Speer T.** Innate immunity in CKD-associated vascular diseases. *Nephrol Dial Transplant* 31: 1813-1821, 2016.
173. **Zhang J, Hua G, Zhang X, Tong R, Du X, and Li Z.** Regulatory T cells/T-helper cell 17 functional imbalance in uraemic patients on maintenance haemodialysis: a pivotal link between microinflammation and adverse cardiovascular events. *Nephrology (Carlton)* 15: 33-41, 2010.
174. **Zhang J, Rane G, Dai X, Shanmugam MK, Arfuso F, Samy RP, Lai MK, Kappei D, Kumar AP, and Sethi G.** Ageing and the telomere connection: An intimate relationship with inflammation. *Ageing Res Rev* 25: 55-69, 2016.
175. **Zhao MM, Xu MJ, Cai Y, Zhao G, Guan Y, Kong W, Tang C, and Wang X.** Mitochondrial reactive oxygen species promote p65 nuclear translocation mediating high-phosphate-induced vascular calcification in vitro and in vivo. *Kidney Int* 79: 1071-1079, 2011.
176. **Zhou T, Zhang M, Zhao L, Li A, and Qin X.** Activation of Nrf2 contributes to the protective effect of Exendin-4 against angiotensin II-induced vascular smooth muscle cell senescence. *Am J Physiol Cell Physiol* 311: C572-C582, 2016.
177. **Zhu N, Yuan W, Zhou Y, Liu J, Bao J, Hao J, and Miao W.** High mobility group box protein-1 correlates with microinflammatory state and nutritional status in continuous ambulatory peritoneal dialysis patients. *J Artif Organs* 14: 125-132, 2011.

971 **Figure 1. Basic mechanisms of uremic inflammation**

972 **Figure 2. Causes of uremic inflammation**

973 **Figure 3. Inflammation concerns and consequences**

974 **Figure 4. Effects of systemic inflammation on homeostasis**

---

<sup>i</sup> In which uremia is defined as the medical condition produced by the toxic effects of abnormally high concentrations of nitrogenous substances in the blood as a result of the kidney's failure to expel waste products by way of the urine (<https://www.britannica.com/science/uremia>)









